Notes

Ehretianone, a Novel Quinonoid Xanthene from *Ehretia buxifolia* with Antisnake Venom Activity

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Ehretianone (1), a new quinonoid xanthene, together with known sterols, was isolated from a MeOH extract of the root bark of *Ehretia buxifolia*. The structure of ehretianone was elucidated as 7-hydroxy-9a α -(3-methylbut-2-enyl)-4a α ,9 α -(2-methylprop-2-enyl)-4a,9a-dihydro-1,4-dioxo-xanthene on the basis of spectroscopic data and X-ray crystallographic analysis. The antisnake venom activity of ehretianone against *Echis carinatus* venom in mice is also reported.

Ehretia buxifolia Roxb. (Boraginaceae) (syn. E. microphylla Lam.) is reported to be useful medicinally in the Ayurvedic and Siddha systems of medicine in India.¹ From an EtOH extract of the aerial parts of the plant bauerenol, ursolic acid and a quinonoid, microphyllone, have been reported.² A MeOH extract of the leaves has afforded rosmarinic acid, bauerenol, α -amyrin, β -amyrin, astragalin, and nicotoflorin.³ A field survey conducted in the Chengalpattu District, Tamilnadu, India, revealed that the root bark of the plant is used by the Irula tribe as an antidote to Echis carinatus envenomation.⁴ The root bark powder is given orally, and it is claimed to stop hemorrhaging caused by the venom. We report herein the isolation and structure elucidation of a new quinonoid xanthene, ehretianone (1), from a MeOH extract of the root bark of the plant. In addition to ehretianone, a sterol mixture containing β -sitosterol, stigmasterol, stigmastanol, α -spinasterol, campesterol, and cholesterol was also obtained.

The antisnake venom activity of ehretianone (1) against *E. carinatus* envenomation in mice also has been investigated.

The MeOH extract of the root bark of *E. buxifolia* was subjected to column chromatography over Si gel. Elution of the column with C_6H_6 -EtOAc (9:1) gave ehretianone (1), which was crystallized from a hexane-ether mixture as red-orange crystals, mp 152 °C. It gave a green color with alcoholic FeCl₃ (phenol) and a pink

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color with alcoholic KOH (quinone). The EIMS showed a molecular ion peak at m/z 350, which analyzed for C₂₂H₂₂O₄. The IR spectrum showed hydroxyl (3350 cm⁻¹), quinonoid carbonyl (1670 cm⁻¹), olefinic double bond, and aromatic (1600, 1590, 1570, 970, 880, 855, 820, and 795 $\mbox{cm}^{-1}\mbox{)}$ absorptions. The UV spectrum showed peaks at 210, 230 (sh), and 295 nm, indicating extended conjugation in the molecule. The ¹H-NMR (CDCl₃ and C₆D₆) and ¹³C-NMR (CDCl₃) spectra (Table 1) revealed the presence of a phenolic hydroxyl ($\delta_{\rm H}$ 4.83, 1H, br s, D₂O exchangeable) and an isopentenyl side chain with two vinylic methyls ($\delta_{\rm H}$ 1.48 and 1.66, 3H each, s; $\delta_{\rm C}$ 17.90 and 25.90), a trisubstituted doublebond proton ($\delta_{\rm H}$ 4.98, 1H, t, J = 7.5 Hz; $\delta_{\rm C}$ 117.08), and a methylene group attached to a saturated tertiary carbon atom ($\delta_{\rm H}$ 2.30 and 2.51, 1H each dd, J = 7.3, 14.6 Hz; $\delta_{\rm C}$ 31.80). Two 1,4-quinone carbonyls appeared as singlets at $\delta_{\rm C}$ 198.53 and 193.29, and a pair of orthocoupled quinone ring protons were apparent ($\delta_{\rm H}$ 6.46

Table 1. ¹H-NMR and ¹³C-NMR Data of Ehretianone (1)

	∂_{H}		
position	CDCl ₃ (mult., <i>J</i> in MHz)	C ₆ D ₆	δ_{C}
1			198.53
2	6.46 (d, 10.7)	5.92	138.10
3	6.80 (d, 10.7)	6.25	138.87
4			193.29
5	6.58 (d, 8.8)	6.67	117.36
6	6.48 (dd, 8.3, 2.9)	6.28	114.31
7			149.85
8	6.54 (d, 2.9)	6.42	114.57
9	3.58 (d, 6.3)	3.63	37.52
OH	4.83 (br s, D ₂ O exchangeable)		
1′	2.30, 2.51 (each dd, 7.3, 14.6)	2.18, 2.42	31.80
2'	4.98 (t, 7.5)	4.98	117.08
3′			131.48
4'	1.66 (s)	1.37	25.90
5'	1.48 (s)	1.33	17.90
1″	2.52, 2.80 (each d, 19.5)	2.62, 2.72	35.78
2″			136.85
3″	5.62 (d, 6.3)	5.38	122.75
4‴	1.74 (s)	1.58	22.60
4a			81.00
8a			127.96
9a			53.93
10a			144.87

Table 2. Cross-Peaks in the ¹H-¹H Long-range COSY NMR

 Spectrum of Ehretianone (1) (non-aromatic region)

¹ Η (δ ppm)	¹ Η (δ ppm)
H-4' (1.66)	H-5' (1.48), H-2' (4.98), H-1' (2.30, 2.51)
H-5' (1.48)	H-4' (1.66), H-2' (4.98), H-1' (2.30, 2.51)
H-1' (2.30)	H-5' (1.48), H-4' (1.66), H-1' gem (2.51), H-2' (4.98)
H-1' (2.51)	H-5' (1.48), H-4' (1.66), H-1' gem (2.30), H-2' (4.98)
H-2' (4.98)	H-4' (1.66), H-5' (1.48), H-1' (2.30, 2.51)
H-4" (1.74)	H-1" (2.52, 2.80), H-3" (5.62), H-9 (3.58)
H-1" (2.52)	H-4" (1.74), H-1" gem (2.80), H-3" (5.62)
H-1" (2.80)	H-4" (1.74), H-1" gem (2.52), H-3" (5.62)
H-3" (5.62)	H-4" (1.74), H-9 (3.58), H-1" (2.52, 2.80)
H-2 (6.46)	H-3 (6.80)
H-3 (6.80)	H-2 (6.46)

and 6.80, 1H each, d, J = 10.7 Hz). The coupling pattern of the three aromatic protons ($\delta_{\rm H}$ 6.48, 6.54, and 6.58) in the ¹H-NMR spectrum showed the presence of a 1,3,4-trisubstituted benzene ring. The spectral data also showed the presence of a methylprop-2-enyl bridge, which is also present in microphyllone previously reported from *E. buxifolia*.²

The ¹H-NMR spectrum of ehretianone (**1**) in C_6D_6 was more highly resolved than when run in CDCl₃, with the overlapping signals well separated and the quinonoid protons moving upfield and being well separated from the aromatic protons. Similarly, the quartet due to H-1", which was found to overlap with the double doublets due to H-1', was also well separated. The ¹³C-NMR spectrum of **1** showed three methyl groups, two methylene groups, eight methine groups, and nine quaternary carbon atoms. The assignments were made by comparison with model compounds.^{5,6}

The ¹H- and ¹³C-NMR assignments of **1** were confirmed by ¹H–¹H long-range and ¹H–¹³C one-bond COSY spectra. The cross-peaks observed between the various protons in the non-aromatic region of the ¹H– ¹H long-range COSY NMR spectrum of ehretianone (**1**) are given in Table 2. The assignments of the quaternary carbon atoms are based on model compounds.

Difficulty in assembling the above groups with the correct stereochemical details led to the study of ehretianone (1) by X-ray crystallography. The structure of



Figure 1. Perspective view of the molecular structure of ehretianone (1).

ehretianone (1) was thereby elucidated as 7-hydroxy- $9a\alpha$ -(3-methylbut-2-enyl)- $4a\alpha$, 9α -(2-methylprop-2-enyl)-4a,9a-dihydro-1,4-dioxoxanthene. The perspective view of the molecule as obtained by the single-crystal X-ray analysis of ehretianone (1) is shown in Figure 1. The pyran ring is puckered, with the total puckering amplitude being 0.664(7) Å. The puckering causes the pyran ring to be distorted into a sofa conformation. The molecule is folded at both the ring junctions C-9a-C-4a and C-8a–C-10a. The quinone ring adopts a twist conformation. The twist runs through the atoms C-2 and C-4a such that the dihedral angle between the two halves of the ring is 28.2(5)°. The bridge connecting C-9 and C-4a is α -linked and adopts a sofa conformation. Final positional parameters and equivalent isotropic temperature factors of non-hydrogen atoms are given in Table 3. In the packing of the molecules, intermolecular hydrogen bonding was absent, and the molecules were held together by van der Waals forces of attraction.

The antisnake venom activity of ehretianone (1) was determined using the LD_{50} of *E. carinatus* venom in mice by the subcutaneous route. The LD_{50} of the venom used was 6.65 mg/kg. The compound did not show any toxic effect up to 100 mg/kg. In vitro neutralization studies showed that compound 1 gave significant protection at a dosage level of 3.75 mg/kg in mice when challenged with the LD_{50} of the venom (Table 4). The prophylactic and curative activities (compound given after venom injection) are given in Table 5. In the prophylactic treatment, compound 1 was administered 30 min before venom injection and the mortality was reduced by 35.0% as compared with controls. In the curative study, the same dosage of the compound gave significant protection up to 5 min after venom injection.

In preliminary studies, the total MeOH extract of the root bark of the plant also showed all three of the above modes of activity (in vitro neutralization, prophylactic activity, and curative activity). Other potential active principles besides ehretianone (1) are yet to be identified.

Table 3. Final Positional Parameters and Equivalent Isotropic Temperature Factors (Å²) of the Non-Hydrogen Atoms with Estimated Standard Deviations in Parentheses

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atom	x/a	<i>y</i> / <i>b</i>	z/c	$B_{\rm eq}$ (Å ²) ^a	
C-1	-0.1709(8)	1.0023(6)	0.3889(2)	3.4(1)	
0-1	-0.2614(6)	1.0521(4)	0.4244(2)	4.4(1)	
C-2	-0.1142(10)	1.0832(6)	0.3417(3)	4.2(2)	
C-3	-0.0252(11)	1.0305(6)	0.3006(3)	4.5(2)	
C-4	0.0326(9)	0.8817(6)	0.3007(2)	3.9(2)	
0-4	0.0766(8)	0.8261(5)	0.2595(2)	5.5(2)	
C-5	0.2952(8)	1.0689(6)	0.4354(2)	3.2(2)	
C-6	0.2868(8)	1.1405(6)	0.4840(2)	3.3(1)	
C-7	0.1713(9)	1.0998(6)	0.5234(2)	3.4(1)	
0-7	0.1544(10)	1.1695(6)	0.5720(2)	4.7(2)	
C-8	0.0586(8)	0.9882(6)	0.5145(2)	3.0(1)	
C-9	-0.0690(8)	0.8078(6)	0.4500(2)	2.7(1)	
0-10	0.1991(5)	0.8835(4)	0.3786(1)	3.3(1)	
C-10a	0.1854(8)	0.9562(5)	0.4271(2)	2.9(1)	
C-4a	0.0426(7)	0.8158(6)	0.3566(2)	2.9(1)	
C-8a	0.0651(8)	0.9160(5)	0.4666(2)	2.8(1)	
C-9a	-0.1223(8)	0.8475(5)	0.3917(2)	3.0(1)	
C-1′	-0.2906(9)	0.7630(7)	0.3741(3)	3.6(2)	
C-2′	-0.3358(10)	0.7630(6)	0.3152(3)	4.4(2)	
C-3′	-0.3837(9)	0.6548(6)	0.2869(3)	4.5(2)	
C-4′	-0.4294(18)	0.6670(12)	0.2272(4)	7.0(4)	
C-5′	-0.4017(18)	0.5141(8)	0.3088(4)	7.3(3)	
C-1″	0.0767(10)	0.6621(6)	0.3545(3)	3.7(2)	
C-2″	0.0686(9)	0.5950(5)	0.4091(3)	3.4(2)	
C-3″	0.0051(8)	0.6616(5)	0.4519(2)	3.2(1)	
C-4″	0.1288(12)	0.4469(7)	0.4126(4)	4.7(2)	

 ${}^{a}B_{\mathrm{eq}} = (8\pi^{2}/3)\Sigma_{i}\Sigma_{j}U_{ij}a_{i}^{*}a_{j}^{*}a_{i}a_{j}.$

Table 4. *In Vitro* Neutralization Effect of Ehretianone (1) against the LD_{50} (6.65 mg/kg) of *E. carinatus* Venom in Mice

dose of 1^a		percentage mortali	ty
(mg/kg)		test	control
1.25	43.3	(p > 0.05)	50.0
2.50	26.7	(p < 0.05)	48.3
3.75	13.3	(p < 0.001)	51.7

 a Ehretianone (1) and the venom were incubated at 37 $^\circ C$ for 30 min before administration.

Table 5. Effect of Ehretianone (1) (3.75 mg/kg) on the Lethality of *E. carinatus* Venom When Administered to Mice at Various Time Intervals, before and after Administering the LD_{50} (6.65 mg/kg) of the Venom

	pe	percentage mortality	
time of administration (min)		test	control
before administration of venom 30	16.7	(<i>p</i> < 0.001)	51.7
after administration of venom			
5	21.7	(<i>p</i> < 0.001)	53.3
30	38.3	(p > 0.05)	55.0
45	45.0	(<i>p</i> > 0.05)	48.3

Experimental Section

General Experimental Procedures. IR spectra were recorded on a Perkin-Elmer 337 grating spectrophotometer using KBr disks. The UV spectra were taken on a Shimadzu 160 A instrument in MeOH. The mass spectrum was recorded on a Shimadzu GC-MS QP 1000A spectrometer at 70 eV. The sterols were analyzed by GC-MS using a Hewlett-Packard 5972 mass selective detector hooked with a Series II plus 5980 gas chromatograph equipped with electronic control flow. A J & W capillary column (30 m × 0.25 mm i.d.) containing 5% diphenylpolysiloxane and 95% dimethylpolysiloxane, of 0.25- μ m film thickness, was used. The temperature program was 80 to 280 °C, 8 °C/min, at an operating voltage of 70 eV. The ¹H-NMR and ¹³C-NMR spectra were recorded in CDCl₃ or C₆D₆ on a JEOL GSX instrument at 400 MHz and 75 MHz, respectively, using TMS as the internal standard. The 2D COSY NMR studies were performed on a Bruker AMX 500 spectrophotometer in CDCl₃. The ¹H-¹H long-range COSY NMR spectrum was recorded with 2048 and 512 data points in the t_2 and t_1 dimensions, respectively, on the reverse detection probe. The data sets were processed with zero filling in t_1 to give a 1 k × 1 k contour map. Magnitude calculation COSY was used. The one-bond ¹H-¹³C COSY NMR spectrum was obtained using a 5-mm broadband probe. The data were accumulated with 2 k and 512 k points in the t_2 and t_1 dimensions, respectively. Zero filling in the t₁ dimension was used during processing to give a $1 \text{ k} \times 1 \text{ k}$ contour map. The experiment was run in the phase-sensitive mode using the TPPI method.7 X-ray analysis was carried out on an Enraf-Nonius CAD-4 diffractometer with the $\omega/2\theta$ scan mode, λ (MoK α) = 0.71069 Å.

Plant Material. The plant material was collected in Chengalpattu District, Tamilnadu, India, during May 1993. It was identified by Dr. S. Usman Ali, Central Research Institute (Siddha), Madras, and a voucher specimen has been deposited in the herbarium of Captain Srinivasa Murti Drug Research Institute for Ayurveda, Madras.

Extraction and Isolation. The shade-dried and coarsely powdered root bark (800 g) was extracted with MeOH by the cold percolation method (48 h). The extract was filtered and concentrated *in vacuo* (yield, 95 g).

Sterol Mixture. The MeOH extract was subjected to column chromatography over Si gel (100–200 mesh). Elution of the column with C_6H_6 –EtOAc (9:1) gave a sterol mixture. The mixture was subjected to GC–MS analysis. The sterols present were: stigmasterol (39.0%), stigmastanol (29.0%), β -sitosterol (19.0%), campesterol (5.15%), α -spinasterol (2.5%), cholesterol (1.5%), and an unidentified sterol (7.0%).

Ehretianone (1). Further elution of the column with the same solvent mixture gave the new quinonoid xanthene, ehretianone (1). The compound (50 mg, yield 0.00625%) was crystallized from hexane-ether as redorange crystals, mp 152 °C. IR (KBr) ν max 3350, 2990, 2940, 2880, 1670, 1660, 1600, 1590, 1570, 1550, 1380, 1350, 1330, 1260, 1250, 1200, 1110, 1100, 970, 880, 855, 820, 795 cm⁻¹; UV (MeOH) λ max 210, 230 (sh), 295 nm; ¹H NMR and ¹³C NMR data, see Table 1; EIMS (70 eV) m/z [M]⁺ 350 (64), 283 (6), 282 (31), 267 (7), 265 (9), 264 (9), 247 (11), 237 (9), 235 (12), 221 (8), 161 (11), 158 (11), 138 (9), 128 (8), 115 (14), 93 (57), 92 (7), 91 (13), 77 (16), 69 (100), 65 (17), 55 (29). Anal. Calcd for C₂₂H₂₂O₄ C, 75.42; H, 6.28. Found: C, 74.77; H, 6.78.

X-ray Crystallography of 1. Suitable crystals of ehretianone (1) were obtained from a CHCl₃–MeOH mixture by slow evaporation. The crystal belonged to the orthorhombic system, space group $P2_12_12_1$ with a = 7.563(1) Å, b = 9.655(1) Å, c = 24.604(1) Å, Z = 4, V = 1796.47(2) Å³, $D_c = 1.307$ g cm⁻³, and μ (MoK α) = 0.08 mm⁻¹. Of the 1675 reflections collected, only 1265 reflections with $I > 3\sigma(I)$ were used for structural analysis. The structure was solved by direct methods and refined by the full matrix-least squares procedure to a final R = 0.044 and Rw = 0.044 where w = 1.2601

 $(\sigma^2 (F) + 0.000822F^2)$. The positions of the hydrogen atoms were identified by difference Fourier methods. Crystallographic calculations were performed on microvax 11 computer by using the SHELXS 86 structuresolving package.

Antisnake Venom Activity of Ehretianone (1). E. carinatus venom was purchased from the Irula Snake Catchers Industrial Cooperative Society, Mahabalipuram, Chengalpattu District, Tamilnadu, India. The LD₅₀ of the venom was determined in Swiss albino mice (20-22 g), six groups, eight in each. The venom was dissolved in 0.9% saline before use and administered by the subcutaneous route. The animals were observed for 6 h, and the number that died within 24 h was recorded. The LD₅₀ was calculated by the method of Miller and Tainter.⁸ Ehretianone (1) was administered as a fine suspension in water containing Tween 80.

The in vitro neutralization activity of ehretianone (1) was determined by incubating the LD₅₀ of *E. carinatus* venom with various doses of the test compound at 37 °C for 30 min before subcutaneous administration to mice.

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